

Rapid and Quantitative Detection of Malarial Antigen for Microfluidic Point-of-Care Diagnostics in the Developing World



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Abstract

We demonstrate a flow-through membrane immunoassay for malaria on a disposable microfluidic card with integrated dry reagent storage

A novel, point-of-care immunoassay format is demonstrated for malarial diagnosis. The assay system consists of a disposable card with integrated dry reagent storage and an external reader that conducts fluidic actuation and assay quantification. Reagents flow through a porous membrane encased in the card's polymeric laminates to conduct a colorimetric sandwich assay on the membrane's surface. The system produces quantitative results with detection limits on the order of ELISA in under 9 minutes.

Global Health Needs

Effective diagnostic strategies for low-resource communities are enabled by stable reagent storage, rapid results, and assay automation

- The settings of low-resource communities place additional requirements on diagnostic tools.
- A lack of refrigeration demands stable reagent storage in high and fluctuating temperatures.
 - Rapid diagnostic results allow health care workers to diagnose and treat disease in a single visit.
 - Inadequate lab facilities and limited health worker training make self-contained and automated assays valuable to obtaining quality diagnoses.



Assay Format Advantages

Reformatting of lateral flow components allows greater assay control, better integration with microfluidic components, and parallel multiplexing

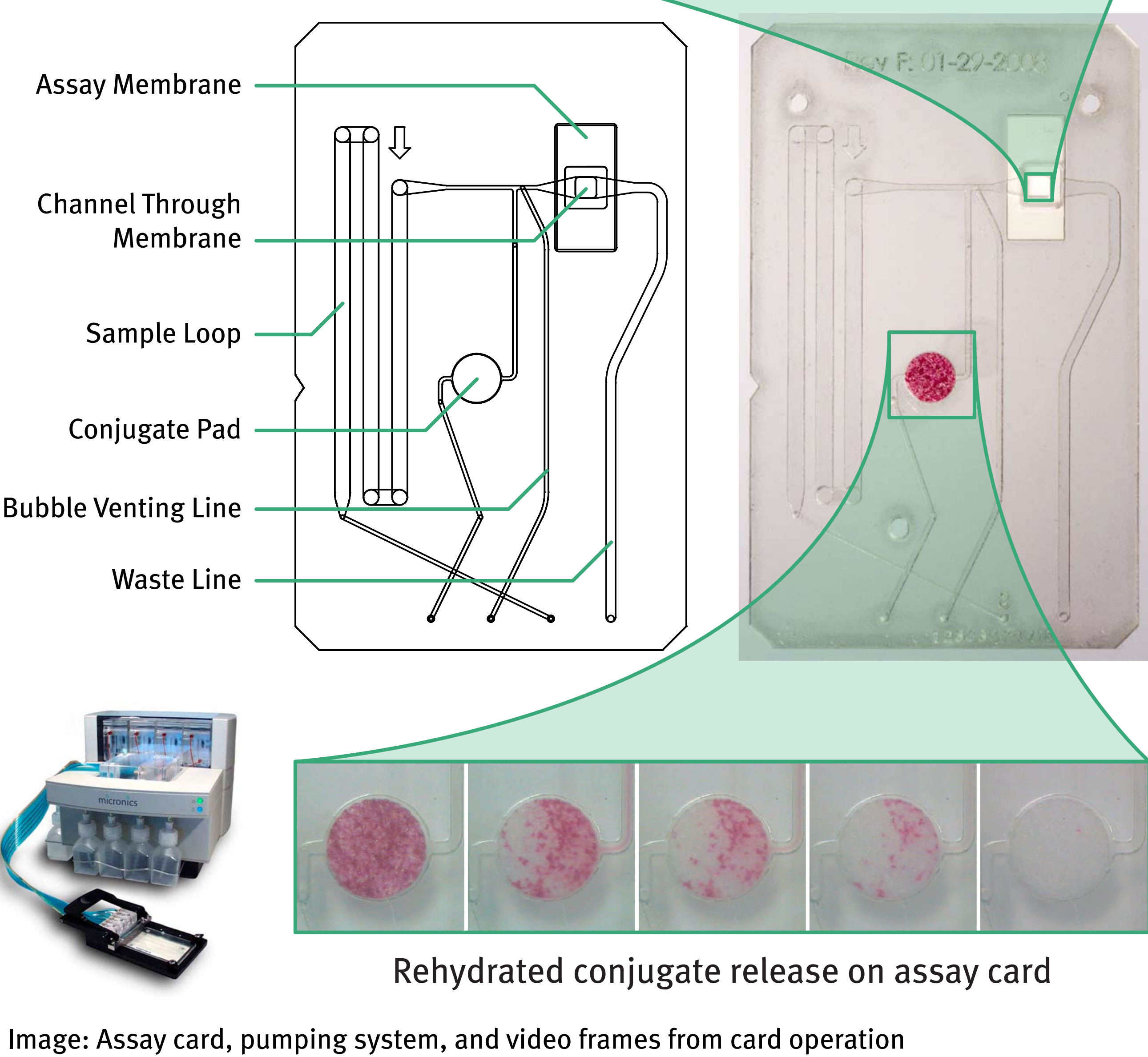
- While sharing some materials with lateral flow assays, this design uses pumps and microfluidic channels to direct fluid through the assay membrane in a transverse direction. This format allows:
- Active control of flow rates
 - Flexible sequencing of reagent addition
 - Simple integration with other microfluidic components
 - Parallel multiplexing by spatial separation of different capture molecules on the membrane.

Additionally, the porous structure of the membrane decreases diffusion distances and provides a high surface area for capture of analyte and label. These benefits have the potential to improve assay signal and generate rapid assay results.

Sample-to-Result Disposable

Using a disposable assay card and an external reader, we integrate dry reagent storage, rapid diagnosis results, and lab-on-a-chip automation in an assay for the malarial biomarker PfHRP2

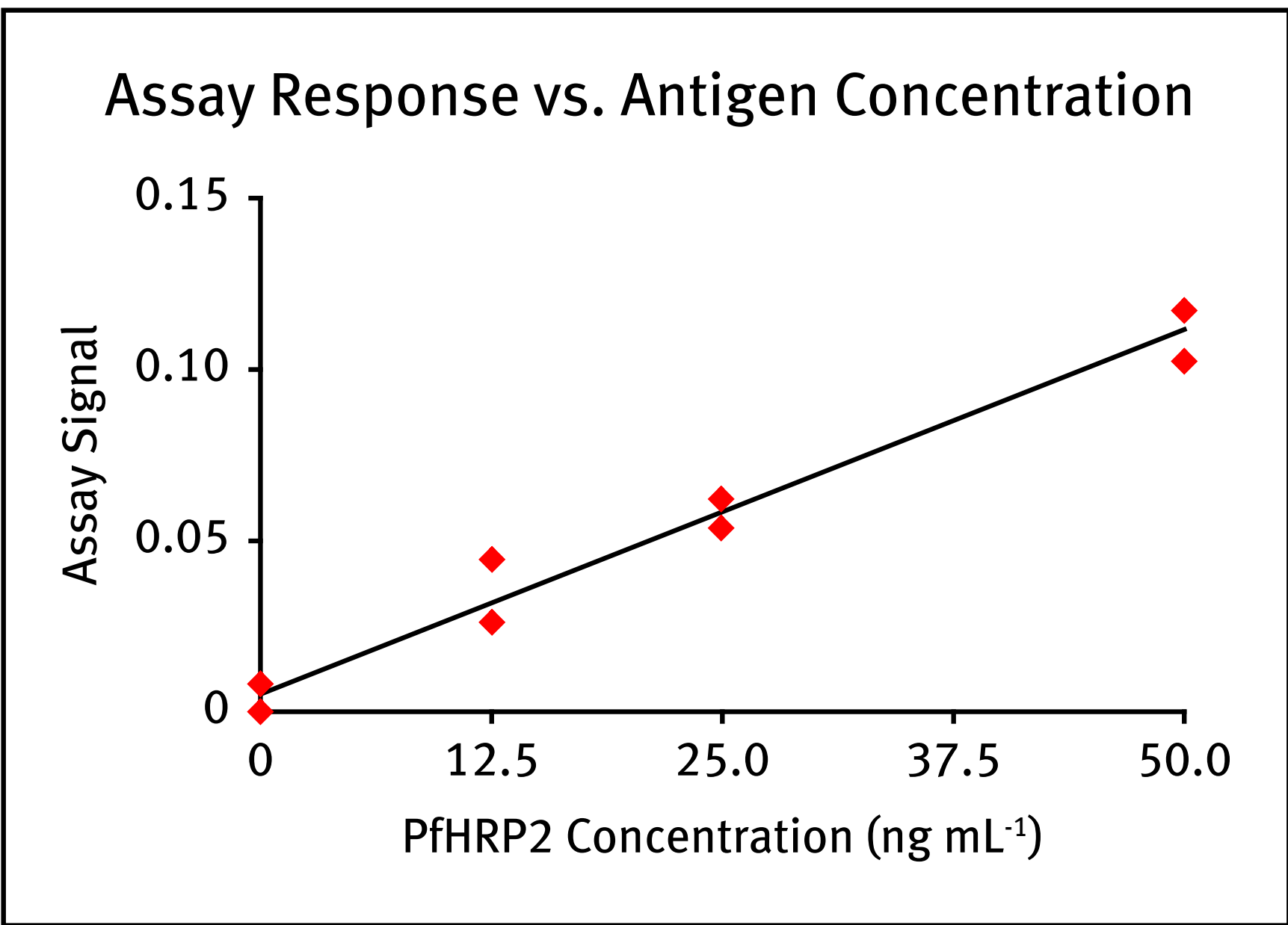
- Assay cards were developed with the following features:
- A porous membrane is embedded in laminate construction to conduct a flow-through membrane immunoassay for PfHRP2, a biomarker for malaria.
 - On-card storage of solid-phase reagent allows unrefrigerated storage of the disposable.
 - A quantitative result in under 9 minutes allows rapid diagnosis of malaria.
 - Assay automation provides reproducible results by limiting human error.
 - A valveless card design is made possible by simultaneous control of fluid lines.



Performance

Assay signal is improved by modifying card geometry and assay timing, giving a detection limit similar to plate-based ELISA (Enzyme-linked immunosorbent assay)

- The assay system was improved in multiple design iterations, demonstrating several characteristics.
- The detection limit for PfHRP2 is comparable to a commercial plate-based ELISA and is performed in a fraction of the time.
 - Assay signals can be increased by lengthening membrane occupancy time of reagents, such as by reducing flow-through areas and flow rates.
 - Taylor dispersion and high density of rehydrated label cause non-uniform exposure of conjugate to the assay membrane. Non-uniform reagent exposure has been addressed by pneumatic pumping approaches and altered protocol timing.



Flow-Through Membrane Immunoassay Format

Forced flow through a porous membrane gives high surface area and allows control of flow rates

- The assay card implements a flow-through membrane immunoassay (FMIA) for PfHRP2 in a sandwich assay format.
- A porous nitrocellulose membrane is patterned with anti-PfHRP2 IgM, and the membrane is held between channels laser-cut in adhesive-backed Mylar laminates.
 - Sample is loaded into the card.
 - Syringe pumps pass the sample through the membrane, and PfHRP2 is captured on the membrane. Buffer washes away unbound sample.
 - Dry anti-PfHRP2 gold-antibody conjugate is rehydrated from a fibrous pad and passed through the membrane. It binds PfHRP2 captured on the membrane, and buffer washes away unbound conjugate.
 - Red spots appear with intensities proportional to the PfHRP2 content of the sample at those membrane locations patterned with capture antibodies.
 - Assay result images are quantified to determine assay signal.

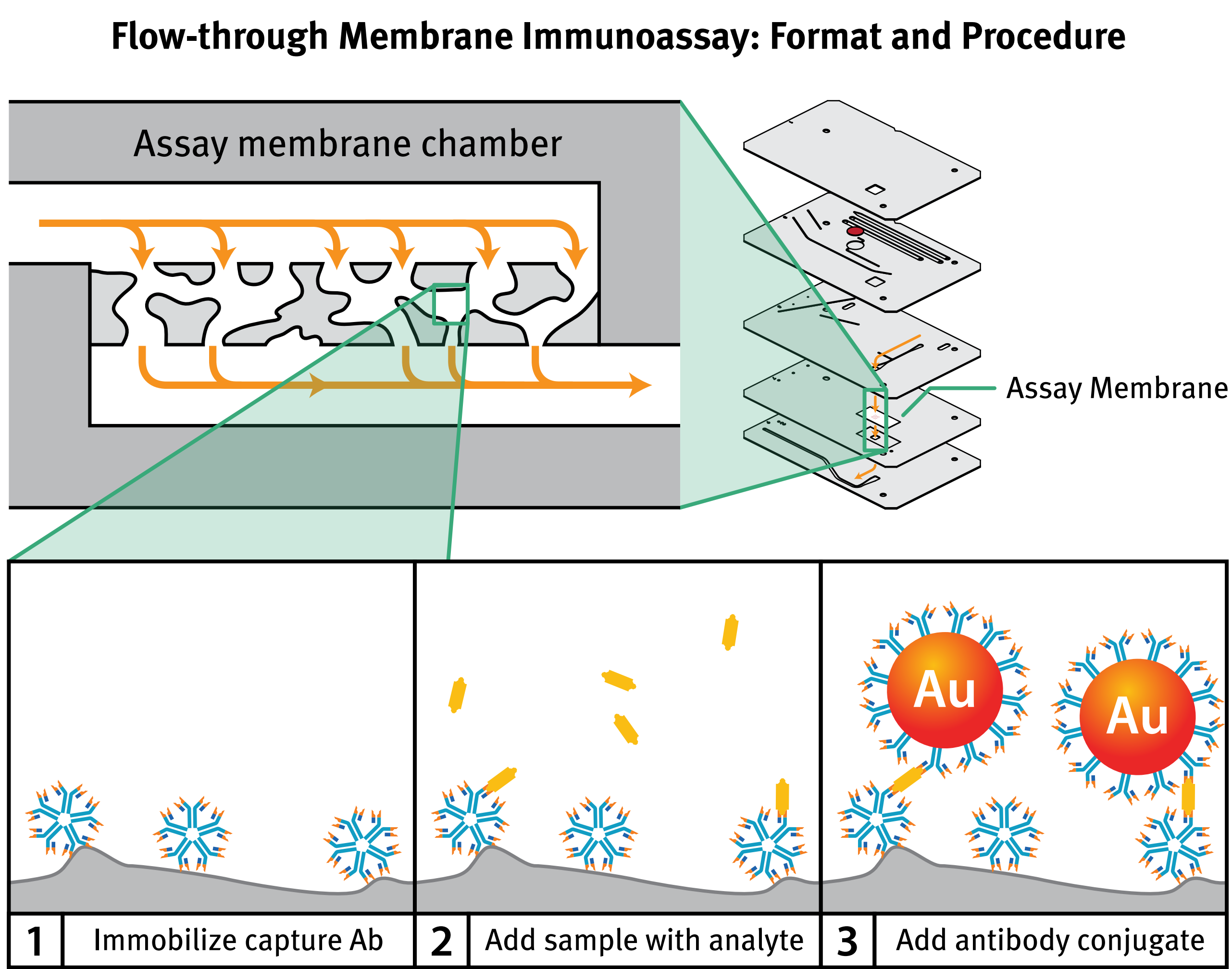


Image: Exploded laminate card, flow-through assay format, and sandwich immunoassay steps

Ongoing Development

Assay multiplexing, pneumatic fluid actuation, and fully-automated assay quantification

- This work provides the basis for ongoing improvements to the assay system demonstrated.
- Assay are multiplexed by patterning multiple capture reagents on the membrane. Diseases diagnosed using the FMIA format include malaria, measles, rickettsia, dengue, and typhoid.
 - Pneumatic fluid actuation simplifies pumping system requirements and reduces Taylor dispersion.
 - Automated image analysis gives repeatable, objective assay quantification.
 - Assay cards are being developed for use with a point-of-care diagnostic reader for the developing world, to include immunoassay and nucleic acid assay results from a finger-stick blood sample.

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